```
(FILE 'CAPLUS' ENTERED AT 12:26:39 ON 30 DEC 2003)
               24 S (PARAMYXOVIRUS (W) (HEMAGGLUTININNEURAMINIDASE OR HEMAGGLUTI
L3
      FILE 'CAPLUS, MEDLINE' ENTERED AT 12:32:55 ON 30 DEC 2003
               24 FILE CAPLUS
L4
               20 FILE MEDLINE
L5
      TOTAL FOR ALL FILES
L6
               44 S L3
L7
               25 DUPLICATE REMOVE L6 (19 DUPLICATES REMOVED)
=> d bib, abs 7-11, 16, 21
     ANSWER 7 OF 25 CAPLUS COPYRIGHT 2003 ACS on STN
L7
      1999:64941 CAPLUS
AN
DN
      130:122133
      Epitopes and active sites of paramyxoviridae proteins and uses thereof
ΤI
      Langedijk, Johannes Petrus Maria; Van Oirschot, Johannes Theodorus
IN
      Stichting Instituut voor Dierhouderij en Diergezondheid, Neth.
PΑ
SO
      PCT Int. Appl., 63 pp.
      CODEN: PIXXD2
DT
      Patent
LA
     English
FAN.CNT 1
                                                   APPLICATION NO. DATE
      PATENT NO.
                         KIND DATE
                         ----
                                -----
                                                   -----
                                                   WO 1998-NL390
PΙ
      WO 9902695
                          A2
                                 19990121
                                                                     19980708
     WO 9902695
                          Α3
                                 19990408
          W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM RW: GH, GM, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
      AU 9882462
                          A1
                                 19990208
                                                  AU 1998-82462
                                                                       19980708
PRAI EP 1997-202100
                           Α
                                 19970708
      WO 1998-NL390
                           W
                                 19980708
AB
      The invention relates to the field of paramyxoviridae, vaccines against
      infections by such viruses, diagnostics for detecting such viruses and
      targets for therapeutics against such viruses. In particular, the
      invention relates to 3-D models identifying a proteinaceous substance
      comprising at least one virus epitope derived from the attachment protein
      of a virus from the family of paramyxoviridae, said epitope corresponding
      to an antigenic site present on the HN protein of paramyxovirus, which
      site is identified as one of loop .beta.1L01, .beta.1L23, .beta.2L01,
      .beta.2L23, .beta.3L01, .beta.3L23, .beta.4L01, .beta.4L23, .beta.5L01,
      .beta.5L23, .beta.6L01 and .beta.6L23, or a functional equiv. thereof.
     Also, the invention relates to a substance blocking the enzymic activity
     of the morbillivirus H protein. As an example, sialic acid was used to
     block the activity of the H protein.
     ANSWER 8 OF 25 CAPLUS COPYRIGHT 2003 ACS on STN
L7
     1999:188573 CAPLUS
AN
ĎΝ
     130:335232
ΤI
     Amino acid substitutions in a conserved region in the stalk of the
     Newcastle disease virus HN glycoprotein spike impair its neuraminidase
     activity in the globular domain
     Wang, Zhiyu; Iorio, Ronald M.
ΑU
CS
     Department of Molecular Genetics and Microbiology, University of
     Massachusetts Medical School, Worcester, MA, 01655-0122, USA
     Journal of General Virology (1999), 80(3), 749-753
SO
```

CODEN: JGVIAY; ISSN: 0022-1317

- PB Society for General Microbiology
- DT Journal
- LA English
- The ectodomain of the paramyxovirus hemagglutininneuraminidase (HN) glycoprotein spike can be divided into two
 regions: a membrane-proximal, stalk-like structure and a terminal globular
 domain. The latter contains all the antibody recognition sites of the
 protein, as well as its receptor recognition and neuraminidase (NA) active
 sites. These two activities of the protein can be sepd. by monoclonal
 antibody functional inhibition studies and mutations in the globular
 domain. Herein, we show that mutation of several conserved residues in
 the stalk of the Newcastle disease virus HN protein markedly decrease its
 NA activity without a significant effect on receptor recognition. Thus,
 mutations in the stalk, distant from the NA active site in the globular
 domain, can also sep. attachment and NA. These results add to an
 increasing body of evidence that the NA activity of this protein is
 dependent on an intact stalk structure.
- RE.CNT 37 THERE ARE 37 CITED REFERENCES AVAILABLE FOR THIS RECORD ALL CITATIONS AVAILABLE IN THE RE FORMAT
- L7 ANSWER 9 OF 25 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 5
- AN 1999:538410 CAPLUS
- DN 131:348236
- TI Anomeric Specificity and Protein-Substrate Interactions Support the 3D Model for the Hemagglutinin-Neuraminidase from Sendai Virus
- AU Bellini, Tiziana; Pasti, Claudia; Manfrinato, Maria Cristina; Tomasi, Maurizio; Dallocchio, Franco
- CS Dipartimento di Biochimica e Biologia Molecolare, Universita di Ferrara, Ferrara, 44100, Italy
- SO Biochemical and Biophysical Research Communications (1999), 262(2), 401-405
 CODEN: BBRCA9; ISSN: 0006-291X
- PB Academic Press
- DT Journal
- LA English

AB

- The 3D structure of paramyxovirus hemagglutininneuraminidase has not yet been resolved; however, a theor. model has been built by using influenza virus and bacterial neuraminidases as template. Two common features of the catalytic mechanism of the neuraminidases of known 3D structure are the anomeric specificity and the involvement of a tyrosine residue in the stabilization of the transition These key features have been investigated on the water-sol. ectodomain of the hemagglutinin-neuraminidase from Sendai virus (cHN). The anomeric specificity of the hydrolysis of the substrate by cHN has been investigated by NMR spectroscopy. The immediate product of the reaction was the .alpha.-anomer, meaning that cHN belongs to glycohydrolases retaining anomeric configuration like influenza virus neuraminidase. Measurements of the UV difference spectrum upon binding of the substrate analog 2,3-dehydro 2-deoxy N-acetyl neuraminic acid indicate the ionization of a tyrosine residue and decreased polarity in the environment of a tryptophan residue. Functional significance of the spectral data was derived from the known structure of influenza neuraminidase, where a tyrosinate ion is involved in the stabilization of the transition-state carbonium ion, and a tryptophan residue is involved in the binding of the acetyl moiety of the substrate. The data give exptl. support to the 3D model of paramyxovirus neuraminidase. (c) 1999 Academic Press.
- RE.CNT 17 THERE ARE 17 CITED REFERENCES AVAILABLE FOR THIS RECORD ALL CITATIONS AVAILABLE IN THE RE FORMAT
- L7 ANSWER 10 OF 25 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 6
- AN 1997:465434 CAPLUS
- DN 127:187297
- TI Sequence and structure alignment of Paramyxoviridae attachment proteins

and discovery of enzymic activity for a morbillivirus hemagglutinin

- AU Langedijk, Johannes P. M.; Daus, Franz J.; van Oirschot, Jan T.
- CS Dep. of Mammalian Virology, Institute for Animal Science and Health, Lelystad, Neth.
- SO Journal of Virology (1997), 71(8), 6155-6167 CODEN: JOVIAM; ISSN: 0022-538X
- PB American Society for Microbiology
- DT Journal
- LA English
- AB On the basis of the conservation of neuraminidase (N) active-site residues in influenza virus N and paramyxovirus hemagglutinin-

neuraminidase (HN), it has been suggested that the three-dimensional (3D) structures of the globular heads of the two proteins are broadly similar. In this study, details of this structural similarity are worked out. Detailed multiple sequence alignment of paramyxovirus HN proteins and influenza virus N proteins was based on the schematic representation of the previously proposed structural similarity. This multiple sequence alignment of paramyxovirus HN proteins was used as an intermediate to align the morbillivirus hemagglutinin (H) proteins with neuraminidase. Hypothetical 3D structures were built for paramyxovirus HN and morbillivirus H, based on homol. modeling. The locations of insertions and deletions, glycosylation sites, active-site residues, and disulfide bridges agree with the proposed 3D structure of HN and H of the Paramyxoviridae. Moreover, details of the modeled H protein predict previously undescribed enzymic activity. This prediction was confirmed for rinderpest virus and peste des petits ruminants virus. The enzymic activity was highly substrate specific, because sialic acid was released only from crude mucins isolated from bovine submaxillary glands. enzymic activity may indicate a general infection mechanism for respiratory viruses, and the active site may prove to be a new target for antiviral compds.

- L7 ANSWER 11 OF 25 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 7
- AN 1997:729261 CAPLUS
- DN 128:58937
- TI Modeling the paramyxovirus hemagglutininneuraminidase protein
- AU Epa, V. Chandana
- CS Biomolecular Research Institute, Parkville, 3052, Australia
- SO Proteins: Structure, Function, and Genetics (1997), 29(3), 264-281 CODEN: PSFGEY; ISSN: 0887-3585
- PB Wiley-Liss
- DT Journal
- LA English
- AB The paramyxovirus hemagglutinin-neuraminidase

(HN) protein exhibits neuraminidase activity and has an active site functionally similar to that in influenza neuraminidases. Earlier work identified conserved amino acids among HN sequences and proposed similarity between HN and influenza neuraminidase sequences. In this work we identify the three-dimensional fold and develop a more detailed model for the HN protein, in the process we examine a variety of protein structure prediction methods. We use the known structures of viral and bacterial neuraminidases as controls in testing the success of protein structure prediction and modeling methods, including knowledge-based threading, discrete three-dimensional environmental profiles, hidden Markov models, neural network secondary structure prediction, pattern matching, and hydropathy plots. The results from threading show that the HN protein sequence has a 6 .beta.-sheet propellor fold and enable us to assign the locations of the individual .beta.-strands. three-dimensional environmental profile and hidden Markov model methods were not successful in this work. The model developed in this work helps to understand better the biol. function of the HN protein and design inhibitors of the enzyme and serves as an assessment of some protein structure prediction methods, esp. after the x-ray crystallog. soln. of

its structure.

RE.CNT 44 THERE ARE 44 CITED REFERENCES AVAILABLE FOR THIS RECORD ALL CITATIONS AVAILABLE IN THE RE FORMAT

- L7 ANSWER 16 OF 25 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 11
- AN 1994:506183 CAPLUS
- DN 121:106183
- TI Site-directed mutagenesis of a conserved hexapeptide in the paramyxovirus hemagglutinin-neuraminidase glycoprotein: effects on antigenic structure and function
- AU Mirza, Anne M.; Deng, Ruitang; Iorio, Ronald M.
- CS Medical School, University of Massachusetts, Worcester, MA, 01655, USA
- SO Journal of Virology (1994), 68(8), 5093-9 CODEN: JOVIAM; ISSN: 0022-538X
- DT Journal
- LA English
- The sequence NRKSCS constitutes the longest linear stretch in the amino AB acid sequence of the hemagglutinin-neuraminidase (HN) glycoprotein of the paramyxoviruses that is completely conserved among all viruses in the group. The authors have used site-directed mutagenesis and expression of the mutated HN protein of one member of the group, Newcastle disease virus, to explore the role of this highly conserved sequence in the structure and function of the protein. Any substitution introduced for each of 4 residues in the sequence, N-234, R-235, K-236, or S-237, results in a drastic decrease in neuraminidase activity relative to that of the wild-type protein. Only substitutions for the terminal serine residue in the sequence had comparatively little effect on this activity. These findings are consistent with prior computer-based predictions of protein secondary structure which had suggested that this domain corresponds to one in the .beta.-sheet propeller structure of the neuraminidase protein of influenza virus closest to the center of the sialic acid binding site and forms part of the enzyme active site. Four of the substitutions, N-234.fwdarw.Y and K-236.fwdarw.E, .fwdarw.Q, and .fwdarw.S, apparently cause a local alteration in the antigenic structure of the protein. This is evidenced by (i) the diminished recognition of the protein only by monoclonal antibodies thought to bind at the neuraminidase active site, among an extensive panel of conformation-specific antibodies, and (ii) the slower rate of migration in SDS-PAGE for all except the K-236.fwdarw.Q mutation. One of the mutations, K-236.fwdarw.S, completely abolishes the ability of the protein to promote cellular fusion when coexpressed with the fusion protein. The latter cannot be explained by a decrease in the relative hemadsorption activity of the protein and suggests that the globular head of the protein may contribute to this process beyond providing receptor recognition.
- L7 ANSWER 21 OF 25 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 16
- AN 1991:39783 CAPLUS
- DN 114:39783
- TI Folding and oligomerization properties of a soluble and secreted form of the paramyxovirus hemagglutinin-neuraminidase qlycoprotein
- AU Parks, Griffith D.; Lamb, Robert A.
- CS Dep. Biochem., Mol. Biol. Cell Biol., Northwestern Univ., Evanston, IL, 60208-3500, USA
- SO Virology (1990), 178(2), 498-508 CODEN: VIRLAX; ISSN: 0042-6822
- DT Journal
- LA English
- AB The paramyxovirus SV5 hemagglutinin-neuraminidase (HN) glycoprotein (a type II integral membrane protein) was converted into a sol. and secreted form (HN-F) by replacing the HN signal/anchor domain with a hydrophobic domain that can act as a cleavable signal sequence. Approx. 40% of the HN-F synthesized was secreted from cells (t1/2 .apprx. 2.5-3 h). The extracellular HN-F mols. were identified as disulfide-linked dimers and

the majority of the population of mols. were resistant to endoglycosidase H digestion. Examn. of the oligomeric form of the secreted HN-F, by sucrose d. gradient sedimentation, indicated that under conditions where HN was a tetramer, HN-F was found to be a dimer, and no extracellular HN-F monomeric species could be detected. Secreted HN-F was fully reactive with conformation-specific monoclonal antibodies and was enzymically active as shown by HN-F having neuraminidase activity. Examn. of the intracellular HN-F species indicated that HN-F monomers were slowly converted to the disulfide-linked form and that under the sucrose d. gradient sedimentation conditions used the HN-F monomers aggregated. Some of the HN-F monomers were degraded intracellularly. These data are discussed in relationship to the seemingly different folding and oligomerization requirements for the intracellular transport of sol. and membrane-bound forms of a glycoprotein. The sol. and biol. active form of HN may be suitable for further structural and enzymic studies